

#### PRODUCT DATASHEET

# Ready-to-Assay™ C5a Anaphylotoxin Receptor Frozen Cells

**CATALOG NUMBER: HTS017RTA** 

CONTENTS: Pack contains 2 vials of mycoplasma-free cells, 1 ml per vial. Fifty (50) mL of Media Component.

STORAGE: Vials are to be stored in liquid N<sub>2</sub>. Media Component at 4°C (-20°C for prolonged storage).

#### **BACKGROUND**

Ready-to-Assay™ GPCR frozen cells are designed for simple, rapid calcium assays with no requirement for intensive cell culturing. Eurofins Discovery Services has optimized the freezing conditions to provide cells with high viability and functionality post-thaw. The user simply thaws the cells and resuspends them in media, dispenses cell suspension into assay plates and, following overnight recovery, assays for calcium response.

C5a is a proinflammatory peptide generated through activation of the complement system, and is more potent than the other anaphylatoxins, C4a and C3a, in activating peripheral blood leukocytes (Gerard and Gerard, 1994). It can actively participate in the regulation of local cytokine network by stimulating production of proinflammatory cytokines (Buchner *et al.*, 1995; Hsu *et al.*, 1999). The proinflammatory effects of C5a are mediated through binding to a specific 7-TM chemoattractant receptor, C5aR (CD88). Binding of C5a to phagocyte C5aR induces chemotaxis, production of superoxide anions, and release of degradative enzymes. Pharmacologic or genetic disruption of C5a/C5aR interaction reduces sepsis (Riedemann *et al.*, 2003), immune complex-induced lung disease (Shushakova *et al.*, 2002), and Arthrogen-induced arthritis (Grant *et al.*, 2002) in experimental models. Cloned human C5a-expressing cell line is made in the Chem-1 host, which supports high levels of recombinant C5a expression on the cell surface and contains high levels of the promiscuous G protein G $\alpha$ 15 to couple the receptor to the calcium signaling pathway. Thus, the cell line is an ideal tool for screening for agonists, antagonists, and modulators at C5a.

#### **USE RESTRICTIONS**

Please see User Agreement (Label License) for further details. One such restriction is that the contents of the supplied vial(s) are limited to a single use and shall not be propagated and/or re-frozen by licensee.

#### **WARNINGS**

For Research Use Only; Not for Use in Diagnostic Procedures Not for Animal or Human Consumption

#### **GMO**

This product contains genetically modified organisms.

Este producto contiene organismos genéticamente modificados.

Questo prodotto contiene degli organismi geneticamente modificati.

Dieses Produkt enthält genetisch modifizierte Organismen.

Ce produit contient organismes génétiquement des modifiés.

Dit product bevat genetisch gewijzigde organismen.

Tämä tuote sisältää geneettisesti muutettuja organismeja.

Denna produkt innehåller genetiskt ändrade organismer.



#### **APPLICATIONS**

Calcium Flux Assays

#### **APPLICATION DATA**

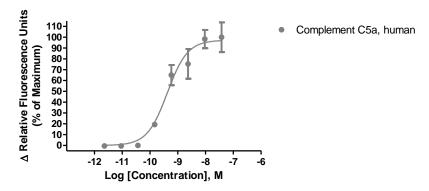


Figure 1. Representative data for activation of C5a receptor. Calcium flux in C5a—expressing Chem-1 cell line induced by human Complement C5a. C5a—expressing Chem-1 cells were loaded with a calcium dye, and calcium flux in response to the indicated ligand(s), 4-fold serial dilution with each concentration performed in duplicate, was determined on a Molecular Devices FLIPR Maximal fluorescence signal obtained in this experiment was 2,000 RLU (Relative Light Units).

Table 1. EC<sub>50</sub> value of C5a-expressing Chem-1 cells.

LIGAND	ASSAY	POTENCY (nM)	REFERENCE
Complement C5a	Calcium Flux	0.4	Eurofins Internal Data

#### **ASSAY SETUP**

- 1. Immediately upon receipt, thaw cells or place cells in liquid nitrogen.
- 2. Thaw cells rapidly by removing from liquid nitrogen and immediately immersing in a 37°C water bath. Immediately after ice has thawed, sterilize the exterior of the vial with 70% ethanol.
- Add 1mL of pre-warmed Media Component to each vial of cells. Place contents from two vials into a 15 mL conical tube and bring the volume to 10 mL of Media Component.
- 4. Centrifuge the cell suspension at 190 x g for four minutes
- 5. Remove supernatant and add 10.5 mL of pre-warmed Media Component to resuspend the cell pellet.
- 6. Seed cell suspension into appropriate assay microplate (100 μL/well for 96-well plate, 25 μL/well for 384-well plate).
- 7. When seeding is complete, place the assay plate at room temperature for 30 minutes.
- 8. Move assay plate to a humidified 37°C 5% CO2 incubator for 24 hours.
- After 24 hour incubation, remove assay plate from the incubator and wash sufficiently with Hank's Balanced Salt Solution (HBSS) supplemented with 20mM HEPES, 2.5mM Probenecid at pH 7.4 to remove all trace of Media Component.



## **Discovery Services**

- 10. Prepare Fluo-8, AM (AAT Bioquest: 21080) Ca<sup>2+</sup> dye by dissolving 1mg of Fluo-8 NW in 200 μL of DMSO. Once dissolved place 10 μL of Fluo-8 NW Ca<sup>2+</sup> dye solution into 10 mL of HBSS 20mM HEPES, 2.5mM Probenecid pH 7.4 buffer and apply to assay microplate (Ca<sup>2+</sup> dye at 10 μL /10 mL is sufficient for loading one (1) microplate).
- 11. Set-up FLIPR to dispense 3x ligand to appropriate wells in the assay plate. Set excitation wavelength at 470-495 nm (FLIPR<sup>TETRA</sup>) or 485 nm (FLIPR1, FLIPR2, FLIPR3) and emission wavelength at 515-565 nm (FLIPR<sup>TETRA</sup>) or emission filter for Ca<sup>2+</sup> dyes (FLIPR1, FLIPR2, FLIPR3). Set pipet tip height to 5 μL below liquid level and dispense rate to 75 μL/sec (96-well format) or 50 μL/sec (384-well format). Set up plate layout and tip layout for each individual experiment. Set time course for 180 seconds, with ligand addition at 10 seconds.
- 12. Ligands are prepared in non-binding surface Corning plates (Corning 3605 96-well or Corning 3574 384-well).
- 13. After the run is complete, negative control correction is applied and data analyzed utilizing the maximum statistic.

#### **ASSAY MATERIALS**

Description	Supplier and Product Number
HBSS	Hyclone: SH30268.02
HEPES 1M Stock	EMD Millipore.: TMS-003-C
Probenicid	Sigma: P8761
Quest Fluo-8™, AM	AAT Bioquest: 21080
Complement C5a ligand	Sigma: C5788
Non-binding white plates (for ligand prep)	Corning: 3605(96-well)/3574(384-well)
Black (clear bottom) tissue-culture treated plates	Corning: 3904(96-well)/3712(384-well)

#### **FLIPR SETTINGS**

Settings for FLIPR<sup>TETRA</sup>® with ICCD camera option

Option	Setting
Read Mode	Fluorescence
Ex/Em	Ex470_495 / Em515_575
Camera Gain	2000
Gate Open	6 %
Exposure Time	0.53
Read Interval	1s
Dispense Volume	50 μl (25 μl for 384-well)
Dispense Height	25 μl (50 μl for 384-well)
Dispense Speed	75 μl L/sec (50 μl for 384-well)
Expel Volume	0 μΙ
Analysis	Subtract Bias Sample 1

#### **HOST CELL**

Chem-1, an adherent rat hematopoietic cell line expressing endogenous  $G\alpha 15$  protein



#### **EXONGENOUS GENE EXPRESSION**

Human C5AR1 cDNA (Accession Number: M62505) expressed from a proprietary proprietary pHS plasmid.

#### RELATED PRODUCTS

PRODUCT NUMBER DESCRIPTION

HTSCHEM-1RTA Ready-to-Assay™ Chem-1 host frozen cells (control cells)
HTS017M ChemiScreen™ C5a Anaphylotoxin receptor membrane prep

#### **REFERENCES**

- 1. Buchner RR, *et al.* (1995) Expression of functional receptors for human C5a anaphylatoxin (CD88) on the human hepatocellular carcinoma cell line HepG2. Stimulation of acute-phase protein-specific mRNA and protein synthesis by human C5a anaphylatoxin. *J. Immunol.* 155: 308.
- 2. Gerard C, Gerard NP. (1994) C5A anaphylatoxin and its seven transmembrane-segment receptor. *Annu.Rev.Immunol.* 12: 775-808.
- 3. Grant, EP, *et al.* (2002) Essential role for the C5a receptor in regulating the effector phase of synovial infiltration and joint destruction in experimental arthritis. *J. Exp. Med.* 196: 1461-1471.
- 4. Hsu, M.H., *et al.* (1999) NF-kappaB activation is required for C5a-induced interleukin-8 gene expression in mononuclear cells. *Blood* 93: 3241-3249.
- 5. Riedemann, NC, et al. (2003) A key role of C5a/C5aR activation for the development of sepsis. J. Leukoc. Biol. 74: 966-970.
- 6. Shushakova, N, *et al.* (2002) C5a anaphylatoxin is a major regulator of activating versus inhibitory FcgammaRs in immune complex-induced lung disease. *J. Clin. Invest.* 110: 1823-30.

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