

#### **PRODUCT DATASHEET**

#### ChemiScreen<sup>™</sup> GAL<sub>1</sub> Receptor Stable Cell Line

#### CATALOG NUMBER: HTS094C

**CONTENTS**: 2 vials of mycoplasma-free cells, 1 mL per vial. **STORAGE**: Vials are to be stored in liquid N<sub>2</sub>.

#### BACKGROUND

ChemiScreen cell lines are constructed in the Chem-1 host, which supports high levels of functional receptor expression on the cell surface. Chem-1 cells contain high endogenous levels of Gα15, a promiscuous G protein, allowing most receptors to couple to the calcium signaling pathway.

Galanin is a 29-30 amino acid peptide originally purified from intestine but later found to be abundant in the CNS. A family of 3 GPCRs, GAL<sub>1</sub>, GAL<sub>2</sub> and GAL<sub>3</sub>, bind to galanin and mediate its biological effects. GAL<sub>1</sub> couples to Gi/o to decrease intracellular cAMP levels (Wang et al., 1998). Galanin and its receptors have been implicated in pain, cognition, seizure activity, depression and drug addiction (Hökfelt, 2005). In particular, GAL<sub>1</sub> mediates the anticonvulsant activity of galanin; mice lacking GAL<sub>1</sub> are affected by spontaneous seizures, and have increased susceptibility to seizures induced by lithium-pilocarpine treatment and electrical perforant path stimulation (McColl et al., 2006; Mazarati et al., 2004). Cloned human GAL<sub>1</sub>-expressing cell line is made in the Chem-1 host, which supports high levels of recombinant GAL<sub>1</sub> expression on the cell surface and contains high levels of the promiscuous G protein G $\alpha$ 15 to couple the receptor to the calcium signaling pathway. Thus, the cell line is an ideal tool for screening for antagonists of interactions between GAL<sub>1</sub> and its ligands.

### **USE RESTRICTIONS**

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#### WARNINGS

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#### GMO

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### **APPLICATIONS**

Calcium Flux Assay

#### **APPLICATION DATA**



Figure 1. Calcium flux in GAL<sub>1</sub>–expressing Chem-1 cell line induced by Galanin. GAL<sub>1</sub>–expressing Chem-1 cells and Wild-Type Chem-1 cells (catalog # HTSCHEM-1) were loaded with Fluo-4 NW and calcium flux in response to recombinant human Galanin (10-6 to 10-10.5 M) was determined in triplicate on a Molecular Devices FLIPR TETRA.

Table 1. EC <sub>50</sub> values of GAL <sub>1</sub> cells.						
	LIGAND	ASSAY	POTENCY EC <sub>50</sub> (nM)	REFERENCE		
	Galanin	Calcium Flux - Fluorescence	1	Eurofins Internal Data		
	* The cell line was te	sted and found to have equivalent	t EC <sub>50</sub> and signal at 1, 3 a	and 6 weeks of continuous culture by		
calcium flux fluorescence.						

### **CELL CULTURE**

Table 2. Recommended Cell Culture Reagents (not provided)

Description	Component	Concentration	Supplier and Product Number
Basal Medium	DMEM high glucose Medium (4.5g/L)	-	Hyclone: SH30022
	Fetal Bovine Serum (FBS)	10%	Hyclone: SH30070.03
	Non-Essential Amino Acids (NEAA)	1X	Hyclone: SH30238.01
	HEPES	1X	EMD Millipore: TMS-003-C
Selection Medium	Basal Medium (see above)	-	
	Geneticin (G418)	250 µg/ml	Invivogen: ant-gn-5
Dissociation	Sterile PBS	-	Hyclone: SH30028.03
	0.25% Trypsin-EDTA	-	Hyclone: SH30042.01
CryoMedium	Basal Medium (see above)	40%	
	Fetal Bovine Serum (FBS)	50%	Hyclone: SH30070.03
	Dimethyl Sulfoxide (DMSO)	10%	Sigma: D2650



### **Cell Handling**

- 1. Upon receipt, directly place cells in liquid nitrogen storage. Consistent cryopreservation is essential for culture integrity.
- 2. Prepare Basal Medium. Prepare 37°C Water Bath. Thaw cells rapidly by removing from liquid nitrogen, and immediately immersing in a 37°C water bath, until 90% thawed. Immediately sterilize the exterior of the vial with 70% ethanol.
- 3. Add vial contents to 15 mL Basal Medium in T75 Tissue Culture Treated Flask. Gently swirl flask and place in a humidified, tissue culture incubator, 37°C, 5% CO<sub>2</sub>.
- 4. 18-24 Hours Post–Thaw, all live cells should be attached. Viability of the cells is expected to be 60-90%, At this time, exchange Basal Medium with Selection Medium.
- 5. When cells are approximately 80% confluent, passage the cells. It is suggested that user expand culture to create >20 vial Master Cell Bank at low passage number. *Cells should be maintained at less than 80% confluency for optimal assay results.*
- 6. Cell Dissociation: Aspirate Culture Medium. Gently wash with 1x Volume PBS. Add 0.1x Volume Warm Trypsin-EDTA. Incubate 4 min, 37°C, until cells dislodge. *If cells do not round up, place in 37°C incubator for additional 2 min.* Neutralize Trypsin and collect cells in 1x Volume Basal Medium.
- 7. Seed Cells for expansion of culture. It is recommended that cell lines are passaged at least once before use in assays.

Table 3. Cell Culture Seeding Suggestions: User should define based on research needs.

Flask Size (cm <sup>2</sup> )	Volume (mL)	Total Cell Number (x10 <sup>6</sup> )	Growth Period (hrs)
T75	15	5.0	24
T75	15	2.0	48
T75	15	0.45	72

## HOST CELL

Chem-1, an adherent cell line expressing the promiscuous G-protein,  $G\alpha 15$ .

#### **EXOGENOUS GENE EXPRESSION**

Full-length human GALR1 cDNA encoding GAL<sub>1</sub> (Accession Number: NM\_001480)

#### **RELATED PRODUCTS**

Product Number	Description
HTSCHEM-1	ChemiScreen <sup>™</sup> Chem-1 Parental Cell Line (control cells)
HTS094RTA	ChemiScreen <sup>™</sup> Receptor GAL <sub>1</sub> Ready to Assay Cells

#### REFERENCES

 Hökfelt T (2005) Galanin and its receptors: Introduction to the Third International Symposium, San Diego, California, USA, 21-22 October 2004. *Neuropeptides* 39: 125-142.



- Mazarati A *et al.* (2004) Patterns of seizures, hippocampal injury and neurogenesis in three models of status epilepticus in galanin receptor type 1 (GalR1) knockout mice. *Neuroscience* 128: 431-41.
- McColl CD *et al.* (2006) Galanin receptor-1 knockout mice exhibit spontaneous epilepsy, abnormal EEGs and altered inhibition in the hippocampus. *Neuropharmacology* 50: 209-18.
- 4. Wang S *et al.* (1998) Differential intracellular signaling of the GalR1 and GalR2 galanin receptor subtypes. *Biochemistry* 37: 6711-7.

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