

DISCOVERY

PRODUCT DATASHEET

ChemiScreen[™] MC₅ Melanocortin Membrane Preparation

CATALOG NUMBER:	HTS155M	QUANTITY:	200 units
LOT NUMBER:	SC20190807	VOLUME/CONCENTRATION:	1 mL, 2 mg/mL

BACKGROUND: The melanocortin system consists of five seven-transmembrane spanning G-protein coupled receptors (MC_1 - MC_5). Among the five members of the melanocortin receptor family, MC_2 and MC_5 are expressed in peripheral tissues. The MC_2 receptor (ACTH receptor) is almost exclusively expressed in the adrenal cortex whereas MC_5 is expressed in several organs including the adrenal cortex. Studies have shown that targeted disruption of the MC5R gene produced mice with a severe defect in water repulsion and thermoregulation caused by decreased production of sebaceous lipids (Chen *et al.*, 1997). Data show a requirement for the MC_5 in multiple exocrine glands for the production of numerous products, indicative of a coordinated system for regulation of exocrine gland function by melanocortin peptides. (Entwistle *et al.*1990). MC_5 membrane preparations are crude membrane preparations made from our proprietary stable recombinant cell lines to ensure high-level of GPCR surface expression; thus, they are ideal HTS tools for screening of agonists and antagonists of MC_5 .

APPLICATIONS: Radioligand Binding Assay



Figure 1. Saturation Binding for MC₅. 10 μ g/well MC₅ Membrane Preparations were incubated with increasing amount of [¹²⁵I]-NDP- α MSH in the absence (total binding, TB) or presence (nonspecific binding, NSB) of greater than 500-fold excess unlabeled NDP- α MSH. Specific binding (SB) was determined by subtracting NSB from TB. Sample data from a representative lot.

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Figure 2. Competition Binding for MC₅. MC₅ Membrane Preparation and Wild-Type Chem-1 membrane preparation (WT; Catalog # HTS000MC1), each at 10 μ g/well, were incubated with 0.3 nM [¹²⁵]-NDP- α MSH and increasing concentrations of unlabeled NDP- α MSH, and more than 15- fold signal:background was obtained. Representative sample data.

SPECIFICATIONS: 1 unit = 10 μg B_{max} for [¹²⁵I]-NDP-αMSH binding: 2.84 pmol/mg K_d for [¹²⁵I]-NDP-αMSH binding: 0.43 nM Signal:background: ≥15-fold

TRANSFECTION: Full-length human MC5R cDNA encoding MC_5 (Accession Number: NM_005913).

HOST CELLS: Chem-1, an adherent mammalian cell line without any endogenous MC₅ expression.

RECOMMENDED ASSAY CONDITIONS: Membranes are mixed with radioactive ligand and unlabeled competitor (see Figures 1 and 2 for concentrations tested) in binding buffer in a nonbinding 96-well plate, and incubated for 1-2 h. Prior to filtration, an FC 96-well harvest plate (EMD Millipore cat. # MAHF C1H) is coated with 0.33% polyethyleneimine for 30 min, then washed with 50 mM HEPES, pH 7.4, 500 mM NaCl. Binding reaction is transferred to the filter plate, and washed 3 times (1 mL per well per wash) with Wash Buffer. The plate is dried and counted.

Binding Buffer: 50 mM HEPES, pH 7.4, 5 mM MgCl₂, 1 mM CaCl₂, filtered and stored at 4°C.

Radioligand: [¹²⁵I]-NDP-αMSH (PerkinElmer#: NEX-352)

Wash Buffer: 50 mM HEPES, pH 7.4, 500 mM NaCl, filtered and stored at 4°C.

One package contains enough membranes for at least 200 assays (units), where a unit is the amount of membrane that will yield greater than 50-fold signal:background with [125 I] NDP- α MSH at 0.3 nM.



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- PRESENTATION:
 Liquid in packaging buffer: 50 mM Tris pH 7.4, 10% glycerol and 1% BSA with no preservatives. Packaging method: Membrane proteins were adjusted to the indicated concentration in 1 ml packaging buffer, rapidly frozen, and stored at -80°C.

 STORAGE/HANDLING:
 Store at -70°C. Product is stable for at least 6 months from the date of receipt when stored as directed. Avoid repeated freeze/thaw cycles.

 REFERENCES:
 1. Chen W et al. (1997). Exocrine gland dysfunction in MC5-R-deficient mice: evidence for
 - coordinated regulation of exocrine gland function by melanocortin peptides. *Cell* 91:789-798.
 - 2. Entwistle ML *et al.* (1990). Characterization of functional melanotropin receptors in lacrimal glands of the rat. *Peptides* 11:477-483.

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