

#### PRODUCT DATASHEET

# Ready-to-Assay™ Secretin Glucagon Receptor Frozen Cells

**CATALOG NUMBER: HTS174RTA** 

CONTENTS: Pack contains 2 vials of mycoplasma-free cells, 1 ml per vial. Fifty (50) mL of Media Component.

STORAGE: Vials are to be stored in liquid N<sub>2</sub>. Media Component at 4°C (-20°C for prolonged storage).

#### **BACKGROUND**

Ready-to-Assay™ GPCR frozen cells are designed for simple, rapid calcium assays with no requirement for intensive cell culturing. Eurofins Discovery Services has optimized the freezing conditions to provide cells with high viability and functionality post-thaw. The user simply thaws the cells and resuspends them in media, dispenses cell suspension into assay plates and, following overnight recovery, assays for calcium response.

Secretin, a member of the secretin-glucagon peptide hormone family, is a 27 amino acid peptide that was originally isolated from the duodenum. In the duodenum, secretin is released to stimulate the release of digestive juices in the pancreas (Bayliss et~al, 1902). The receptor for secretin is a class 2 (or class B) G protein coupled receptor that signals through  $G_s$  to stimulate cAMP production (Dong et~al, 2002). Along with its traditional role in the pancreas, studies in secretin-deficient mice have shown miscommunication between the CA3 Schaffer collateral and CA1 pyramidal neurons, causing a deficiency in synaptic transmission (Nishijima et~al, 2006). This miscommunication of CA1 dendrites is found in Autism, Rett Syndrome, and most forms of mental retardation, suggesting secretin could be a potential target for treatment of these disorders. Cloned human Secretin-expressing cell line is made in the Chem-1 host, which supports high levels of recombinant Secretin expression on the cell surface and contains high levels of the promiscuous G protein Ga15 to couple the receptor to the calcium signaling pathway. Thus, the cell line is an ideal tool for screening for agonists, antagonists and modulators at Secretin.

#### **USE RESTRICTIONS**

Please see User Agreement (Label License) for further details. One such restriction is that the contents of the supplied vial(s) are limited to a single use and shall not be propagated and/or re-frozen by licensee.

#### **WARNINGS**

For Research Use Only; Not for Use in Diagnostic Procedures Not for Animal or Human Consumption

#### **GMC**

This product contains genetically modified organisms.
Este producto contiene organismos genéticamente modificados.
Questo prodotto contiene degli organismi geneticamente modificati.
Dieses Produkt enthält genetisch modifizierte Organismen.
Ce produit contient organismes génétiquement des modifiés.
Dit product bevat genetisch gewijzigde organismen.
Tämä tuote sisältää geneettisesti muutettuja organismeja.
Denna produkt innehåller genetiskt ändrade organismer.



#### **APPLICATIONS**

Calcium Flux Fluorescent Assays, cAMP Accumulation Assays

#### APPLICATION DATA

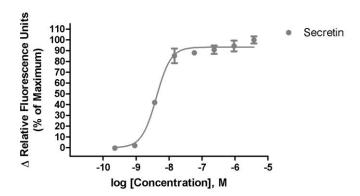


Figure 1. Representative data for activation of Secretin receptor. Calcium flux in Secretin-expressing Chem-1 cell line induced by Secretin. Secretin-expressing Chem-1 cells were loaded with a calcium dye, and calcium flux in response to the indicated ligand(s), 4-fold serial dilution with each concentration performed in duplicate, was determined on a Molecular Devices FLIPR TETRA. Maximal fluorescence signal obtained in this experiment was 3,500 RLU (Relative Light Units).

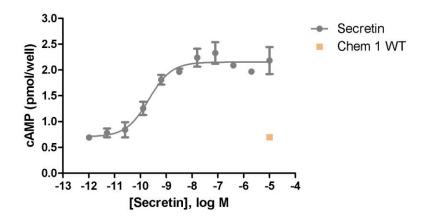


Figure 2. Representative data for activation of Secretin receptor stably expressed in Chem-1 cells induced by Secreting using a cAMP accumulation assay. Secretin—expressing Chem-1 cells were seeded at 100,000 cells per well into a 96-well plate, and the following day the cells were treated with Secretin for 15 minutes in the presence of 2.0 mM IBMX and 0.5% DMSO to determine receptor-mediated cAMP generation using a time-resolved fluorescence resonance energy transfer (TR-FRET) assay measured on the BioTek Synergy. Maximal cAMP response obtained in this experiment was 2.5 pmol/well. Similarly parental cells (catalog #: HTSCHEM-1) were tested to determine the specificity of the resulting signal.



# **Discovery Services**

Table 1. EC<sub>50</sub> values of Secretin-expressing Chem-1 cells.

LIGAND	ASSAY	POTENCY (nM)	REFERENCE
Secretin	Calcium Flux	4	Eurofins Internal Data
Secretin	cAMP Accumulation	0.2	Eurofins Internal Data

#### **ASSAY SETUP**

- Immediately upon receipt, thaw cells or place cells in liquid nitrogen.
- 2. Thaw cells rapidly by removing from liquid nitrogen and immediately immersing in a 37°C water bath. Immediately after ice has thawed, sterilize the exterior of the vial with 70% ethanol.
- Add 1mL of pre-warmed Media Component to each vial of cells. Place contents from two vials into a 15 mL conical tube and bring the volume to 10 mL of Media Component.
- 4. Centrifuge the cell suspension at 190 x g for four minutes
- Remove supernatant and add 10.5 mL of pre-warmed Media Component to resuspend the cell pellet.
- 6. Seed cell suspension into appropriate assay microplate (100 μL/well for 96-well plate, 25 μL/well for 384-well plate).
- 7. When seeding is complete, place the assay plate at room temperature for 30 minutes.
- 8. Move assay plate to a humidified 37°C 5% CO2 incubator for 24 hours.
- After 24 hour incubation, remove assay plate from the incubator and wash sufficiently with Hank's Balanced Salt Solution (HBSS) supplemented with 20mM HEPES, 2.5mM Probenecid at pH 7.4 to remove all trace of Media Component.
- 10. Prepare Fluo-8, AM (AAT Bioquest: 21080) Ca<sup>2+</sup> dye by dissolving 1mg of Fluo-8 NW in 200 μL of DMSO. Once dissolved place 10 μL of Fluo-8 NW Ca<sup>2+</sup> dye solution into 10 mL of HBSS 20mM HEPES, 2.5mM Probenecid pH 7.4 buffer and apply to assay microplate (Ca<sup>2+</sup> dye at 10 μL /10 mL is sufficient for loading one (1) microplate).
- 11. Set-up FLIPR to dispense 3x ligand to appropriate wells in the assay plate. Set excitation wavelength at 470-495 nm (FLIPR<sup>TETRA</sup>) or 485 nm (FLIPR1, FLIPR2, FLIPR3) and emission wavelength at 515-565 nm (FLIPR<sup>TETRA</sup>) or emission filter for Ca<sup>2+</sup> dyes (FLIPR1, FLIPR2, FLIPR3). Set pipet tip height to 5 μL below liquid level and dispense rate to 75 μL/sec (96-well format) or 50 μL/sec (384-well format). Set up plate layout and tip layout for each individual experiment. Set time course for 180 seconds, with ligand addition at 10 seconds.
- 12. Ligands are prepared in non-binding surface Corning plates (Corning 3605 96-well or Corning 3574 384-well).
- 13. After the run is complete, negative control correction is applied and data analyzed utilizing the maximum statistic.

#### **ASSAY MATERIALS**

Description	Supplier and Product Number
HBSS	Hyclone: SH30268.02
HEPES 1M Stock	EMD Millipore.: TMS-003-C
Probenicid	Sigma: P8761
Quest Fluo-8™, AM	AAT Bioquest: 21080
Secretin ligand	Tocris: 1918
Non-binding white plates (for ligand prep)	Corning: 3605(96-well)/3574(384-well)
Black (clear bottom) tissue-culture treated plates	Corning: 3904(96-well)/3712(384-well)

#### **FLIPR SETTINGS**

Settings for FLIPR<sup>TETRA</sup>® with ICCD camera option

Option	Setting
Read Mode	Fluorescence
Ex/Em	Ex470_495 / Em515_575
Camera Gain	2000
Gate Open	6 %
Exposure Time	0.53
Read Interval	1s
Dispense Volume	50 μl (25 μl for 384-well)
Dispense Height	25 µl (50 µl for 384-well)
Dispense Speed	75 μl L/sec (50 μl for 384-well)
Expel Volume	0 μΙ
Analysis	Subtract Bias Sample 1

#### **HOST CELL**

Chem-1, an adherent rat hematopoietic cell line expressing endogenous  $G\alpha 15$  protein.

#### **EXONGENOUS GENE EXPRESSION**

SCTR cDNA (Accession Number: NM\_002980; see CODING SEQUENCE below) expressed from a proprietary expressed from a proprietary pHS plasmid.

#### **CODING SEQUENCE**

1	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	90 30
91 31	CTATGTGACGTGCTACAAGTGCTGTGGGAAGACAAGACCAGTGCCTGCAGGAACTCTCCAGAGAGCAGAACAGGAGACCTGGGCACGAACL C D V L Q V L W E E Q D Q C L Q E L S R E Q T G D L G T E	180 60
181 61	CAGCCAGTGCCAGGTTGTGAGGGGATGTGGGACAACATAAGCTGCTGGCCCTCTTCTGTGCCGGGCCGGATGGTGGAGGTGGAATGCCCCQQPVPGGCCGGGCCGGATGGTGGAGTGGAATGCCCCQPVPGGCCGGGCCGGATGGTGGAGTGGAATGCCCCQQPVPGGAGGTGGAATGCCCCQQPVPGGAGGTGGAATGCCCCQQPVPGGAGGTGGAATGCCCCQQPVPGAGGTGGAATGCCCCQQPVPGAGGAGGTGGAATGCCCCQQPVPGAGGTGGAATGCCCCQQPVPGAGGTGGAATGCCCCCCTCTTCTGTGCCGGGCCGGATGGTGGAGGTGGAATGCCCCCCCTCTTCTGTGCCGGGCCGGATGGTGGAGGTGGAATGCCCCCCCC	270 90
271 91	$\label{eq:constraints} \begin{array}{cccccccccccccccccccccccccccccccccccc$	360 120
361 121	CTGGCCTGTGGCGTTAATGTGAACGACTCTTCCAACGAGAAGCGGCACTCCTACCTGCTGAAGCTGAAAGTCATGTACACCGTGGGCTACL A C G V N V N D S S N E K R H S Y L L K L K V M Y T V G Y	450 150
451 151	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	540 180
541 181	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	630 210
631 211	CACAGGGCGGCTGCAAGCTGGTCATGGTGCTGTTCCAGTACTGCATCATGGCCAACTACTCCTGGCTGCTGGTGGAAGGCCTCTACCTTH R A G C K L V M V L F Q Y C I M A N Y S W L L V E G L Y L	720 240
721 241	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	810 270
811 271	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	900 300
901 301	CCTGTGATCCTCCATCCTGATTAATTTCATCCTTTTCATAAACATTCTAAGAATCCTGATGAGAAAACTTAGAACCCAAGAAACAAGAP $P$ V I L S I L I N F I L F I N I L R I L M R K L R T Q E T R	990 330
991 331	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1080 360
1081	${\tt TCCCCAGAGGACGCTATGGAGATCCAGCTGTTTTTTGAACTAGCCCTTGGCTCATTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGCCGTCCTCTACTGCTTCCAGGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTACTGCTCAGGACTGGTGGCCGTCCTCTACTGCTTCAGGACTGGTGGCCGTCCTCTAGGCTCAGGACTGGTGGCCGTCCTCTACTGCTCAGGACTGGTGGCCGTCCTCTAGGCTCAGGACTGGTGGCCGTCCTCTAGGCTCAGGACTGGTGGCCGTCCTCTAGGCTCAGGACTGGTGGCCGTCCTCTAGGCTCAGGACTGGTGGCCGTCCTCTAGGCCCTCTCAGGACTGGTGGCCGTCCTCTAGGCCGTCAGGACTGGTGGCCGTCCTCTAGGACTAGGACTGGTGGCCGTCAGGACTGGTGGCCGTCAGGACTGGTGGCCGTCCTAGGACTGGCTGG$	: 1170



# **Discovery Services**

361	S	P	Ε	D	A	M	Ε	I	Q	L	F	F	Ε	L	А	L	G	S	F	Q	G	L	V	V	A	V	L	Y	С	F	390
1171	CTC	AAC	GGG	GAG	GTG	CAG	CTG	GAG	GTT	CAG.	AAG	AAG	TGG	CAG	CAA	TGG	CAC	CTC	CGT	GAG	TTC	CCA	CTC	CAC	ccc	CGT	GC	CTC	CTT	CAGC	1260
391	L	N	G	E	V	Q	L	Ε	V	Q	K	K	M	Q	Q	W	Н	L	R	Ε	F	Ρ	L	Н	P	V	Α	S	F	S	420
1261	1261 AACAGCACCAAGGCCACTTGGAGCAGAGCCAGGGCACCTGCAGGACCAGCATCATCTGA																														
421	N	S	Т	K	A	S	Η	L	E	Q	S	Q	G	T	C	R	T	S	I	I	Str	)									

#### RELATED PRODUCTS

PRODUCT NUMBER DESCRIPTION

HTSCHEM-1RTA Ready-to-Assay™ Chem-1 host frozen cells (control cells)

#### **REFERENCES**

- 1. Bayliss W. et al. (1902) The mechanism of pancreatic secretion. J. Physiol. (Lond) 28:325–353.
- 2. Dong M et al. (2002) Molecular pharmacology of the secretin receptor. Receptors Channels 8:189–200.
- 3. Nishijima I *et al.* (2006) Secretin receptor-deficient mice exhibit impaired synaptic plasticity and social behavior. *Hum Mol Genet* 15(21):3241-50.

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## **Discovery Services**

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