

PRODUCT DATASHEET

ChemiScreen™ P2Y₁₁ Purinergic Receptor Stable Cell Line

CATALOG NUMBER: HTS213C

CONTENTS: 2 vials of mycoplasma-free cells, 1 mL per vial.

STORAGE: Vials are to be stored in liquid N₂.

BACKGROUND

ChemiScreen cell lines are constructed in the 1321N1 host, which supports high levels of functional receptor expression on the cell surface. 1321N1 cells contain high endogenous levels of G_q, a promiscuous G protein, allowing most receptors to couple to the calcium signaling pathway.

The P2Y GPCRs serve as receptors for extracellular nucleotides (Abbracchio et al., 2006). P2Y₁₁ is a relatively low affinity receptor for ATP present humans but not rodents. It is unique among P2Y receptors in increasing both cAMP and calcium levels (Qi et al., 2001). P2Y₁₁ is expressed in immune cells, and it mediates dendritic cell maturation induced by ATP (Wilkin et al., 2001). In addition to ATP, NAD⁺ and NAADP⁺ bind and activate P2Y₁₁ in granulocytes (Moreschi et al., 2006, 2007). An alternate form of P2Y₁₁, in which the second exon of P2Y₁₁ undergoes intergenic splicing with exon 12 of the adjacent SSF1 gene to generate a chimeric protein, is widely expressed in human tissues (Communi et al., 2001). The cloned human P2Y₁₁-expressing cell line is made in the 1321N1 host, which supports high levels of recombinant P2Y₁₁ expression on the cell surface. Thus, the cell line is an ideal tool for screening for agonists and antagonists of P2Y₁₁.

USE RESTRICTIONS

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WARNINGS

For Research Use Only; Not for Use in Diagnostic Procedures
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GMO

This product contains genetically modified organisms.
Este producto contiene organismos genéticamente modificados.
Questo prodotto contiene degli organismi geneticamente modificati.
Dieses Produkt enthält genetisch modifizierte Organismen.
Ce produit contient organismes génétiquement des modifiés.
Dit product bevat genetisch gewijzigde organismen.
Tämä tuote sisältää geneettisesti muutettuja organismeja.
Denna produkt innehåller genetiskt ändrade organismer.

APPLICATIONS

Calcium Flux Fluorescence Assay

APPLICATION DATA

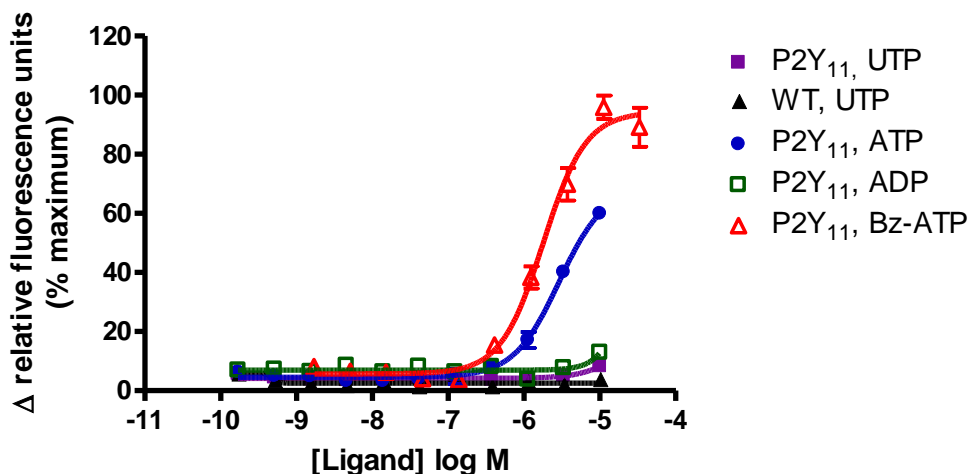


Figure 1. Representative data for activation of the P2Y₁₁ receptor stably expressed in 1321N1 cells induced by UTP, ATP, ADP, and Benzoylbenzoyl-ATP (Bz-ATP) using a fluorescent calcium flux assay. P2Y₁₁-expressing 1321N1 cells were seeded at 50,000 cells per well into a 96-well plate, and the following day the cells were loaded with a fluorescent calcium indicator. Calcium flux in response to the indicated ligand with a final concentration of 0.5% DMSO was determined on a Molecular Devices FLIPR^{TETRA}® with ICCD camera.

Table 1. EC₅₀ values of P2Y₁₁-expressing 1321N1 cells.

LIGAND	ASSAY	POTENCY EC ₅₀ (nM)	REFERENCE
Bz-ATP	Calcium Flux - Fluorescence	2100	Eurofins Internal Data
ATP	Calcium Flux - Fluorescence	5100	Eurofins Internal Data

* The cell line was tested and found to have equivalent EC₅₀ and signal at 1, 3 and 6 weeks of continuous culture by calcium flux fluorescence.

CELL CULTURE

Table 2. Recommended Cell Culture Reagents (not provided)

Description	Component	Concentration	Supplier and Product Number
Basal Medium	DMEM high glucose Medium (4.5g/L)	-	Hyclone: SH30022
	Fetal Bovine Serum (FBS)	10%	Hyclone: SH30070.03
	Non-Essential Amino Acids (NEAA)	1X	Hyclone: SH30238.01
	HEPES	10mM	Millipore Sigma: H3537
	Selection Medium	Basal Medium (see above)	-
Geneticin (G418)		250 µg/ml	Gibco:10131-027
Dissociation	Sterile PBS	-	Hyclone: SH30028.03
	0.25% Trypsin-EDTA	-	Hyclone: SH30042.01
CryoMedium	Basal Medium (see above)	40%	

Fetal Bovine Serum (FBS)	50%	Hyclone: SH30070.03
Dimethyl Sulfoxide (DMSO)	10%	Sigma: D2650

Cell Handling

1. Upon receipt, directly place cells in liquid nitrogen storage. Consistent cryopreservation is essential for culture integrity.
2. Prepare Basal Medium. Prepare 37°C Water Bath. Thaw cells rapidly by removing from liquid nitrogen, and immediately immersing in a 37°C water bath, until 90% thawed. Immediately sterilize the exterior of the vial with 70% ethanol.
3. Add vial contents to 15 mL Basal Medium in T75 Tissue Culture Treated Flask. Gently swirl flask and place in a humidified, tissue culture incubator, 37°C, 5% CO₂.
4. 18-24 Hours Post–Thaw, all live cells should be attached. Viability of the cells is expected to be 60-90%. At this time, exchange Basal Medium with Selection Medium.
5. When cells are approximately 80% confluent, passage the cells. It is suggested that user expand culture to create >20 vial Master Cell Bank at low passage number. *Cells should be maintained at less than 80% confluency for optimal assay results.*
6. Cell Dissociation: Aspirate Culture Medium. Gently wash with 1x Volume PBS. Add 0.1x Volume Warm Trypsin-EDTA. Incubate 4 min, 37°C, until cells dislodge. *If cells do not round up, place in 37° C incubator for additional 2 min.* Neutralize Trypsin and collect cells in 1x Volume Basal Medium.
7. Seed Cells for expansion of culture. It is recommended that cell lines are passaged at least once before use in assays.

Table 3. Cell Culture Seeding Suggestions: *User should define based on research needs.*

Flask Size (cm ²)	Volume (mL)	Total Cell Number (x10 ⁶)	Growth Period (hrs)
T75	15	5.0	24
T75	15	2.0	48
T75	15	0.45	72

ASSAY SETUP

Fluorescence

Table 4. Settings for FLIPR^{TETRA}® with ICCD camera option

Option	Setting
Read Mode	Fluorescence
Ex/Em	Ex470_495 / Em515_575
Camera Gain	2000
Gate Open	6 %
Exposure Time	0.53
Read Interval	1s
Dispense Volume	50 µl (25 µl for 384-well)
Dispense Height	95 µl (50 µl for 384-well)
Dispense Speed	50 µl/sec
Expel Volume	0 µl
Analysis	Subtract Bias Sample 1

Table 5. Assay Materials (Not provided)

Description	Supplier and Product Number
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HBSS	Invitrogen: 14025
HEPES 1M Stock	EMD Millipore: TMS-003-C
Probenicid	Sigma: P8761
Quest Fluo-8™, AM	AAT Bioquest: 21080
ATP	Sigma: A6419
Non-Binding 96/384 well Plates (for ligand prep)	Corning: 3605/ 3574
Black (clear Bottom) cell assay plates	Corning: 3904/ 3712

Assay Protocol – Fluorescence

1. Dissociate Culture as Recommended. Collect in Basal Medium. Document Cell Count and Viability
2. Centrifuge the cell suspension at 190 x g for six min
3. Remove supernatant. Gently resuspend the cell pellet in Basal Medium. *It is suggested that end user optimize cell plating based on individual formats.* (Default: Resuspend in volume to achieve 5×10^5 cells/ml (i.e, if collected 5×10^6 TC, $\frac{5 \times 10^6}{5 \times 10^5/ml} = 10$ mL volume)
4. Seed cell suspension into black, clear bottom plate (100 μ L/well for 96-well plate). *When seeding is complete, place the assay plate at room temperature for 30 min.*
5. Move assay plate to a humidified 37°C 5% CO₂ incubator for 18-24 h.
6. Next day, prepare Assay buffer (HBSS, 20mM HEPES, 2.5 mM Probenicid, pH 7.4) and Loading buffer (Assay buffer with 5 mM Fluo8 Dye). *Note: Please prepare Fluo8 stock according to Manufacturer's Recommendations*
7. Remove medium from assay plate by quickly inverting plate on an absorbent pad and blotting to remove all Media Component.
8. Add Loading buffer to assay plate (100 μ L/well for 96-well plate). Incubate plate for 1.5 h at room temperature, protected from light.
9. Prepare ligands in assay buffer at 3x final concentration in non-binding plates. Use Buffer Only Control Wells for Background Subtraction.
10. Create protocol for ligand addition. Please refer to FLIPR™^{TETRA}® settings provided in Table 2. Set time course for 180 s, with ligand addition at 10 s.
11. After the run is complete, apply subtract bias on sample 1. We recommend using Negative Control Correction with Buffer Only Wells. Export data to according to research needs. For most Calcium Flux analysis using Export of Max Signal to end of run is sufficient.

HOST CELL

1321N1, an adherent cell line lacking the endogenous expression of receptors for P2Y11.

EXOGENOUS GENE EXPRESSION

Full-length human P2RY11 cDNA encoding P2Y₁₁ (Accession Number: NM_002566) and promiscuous G_q protein are expressed in a bicistronic vector

RELATED PRODUCTS

Product Number	Description
HTS211C	ChemiScreen™ P2Y ₄ Purinergic Receptor Stable Cell Line

REFERENCES

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coupled nucleotide receptors: from molecular mechanisms and pathophysiology to therapy. *Pharmacol. Rev.* 58: 281-341.

2. Communi D et al. (1997) Cloning of a human purinergic P2Y receptor coupled to phospholipase C and adenylyl cyclase. *J. Biol. Chem.* 272: 31969-31973.
3. Communi D et al. (1999) Pharmacological characterization of the human P2Y11 receptor. *Br. J. Pharmacol.* 128: 1199-1206.
4. Communi D et al. (2001) Cotranscription and intergenic splicing of human P2Y11 and SSF1 genes. *J. Biol. Chem.* 276:16561-16566.
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7. Qi A.-D. et al. (2001) Differential coupling of the human P2Y11 receptor to phospholipase C and adenylyl cyclase. *Br. J. Pharmacol.* 132: 318-326.
8. Wilkin F et al. (2001) The P2Y11 receptor mediates the ATP-induced maturation of human monocyte-derived dendritic cells. *J. Immunol.* 166: 7172-7177.

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